# Microbial and Molecular Ecology

The INL is expanding the classical methods for identifying important microbial communities and for determining their numbers, activities, and capabilities. By understanding environmental microbial communities and their physiologies, we can use them for a range of environmental restoration, natural resource recovery, and energy development applications. Molecular biology and surface chemistry disciplines are joined to develop new ecological understanding of single-cell resources in both surface and subsurface environments. With the expansion of microbial genome sequencing, INL research will lead to better understanding and application of microbial processes through detailed knowledge of gene expression under different conditions.

### *Progress*

INL findings indicate the value of both traditional culture-based and molecular nonculture-based methods of characterizing microbial communities. Traditional methods yield isolates from unique environments and allow complete physiological characterization such as performed on samples from gas hydrate-rich sediments and deep sandstones and shales of hydrocarbon-rich sedimentary deposits. INL molecular ecology methods have led to assays that identify specific microorganisms that prefer to be attached (versus unattached) in the subsurface; diversity of methanotrophs in a basalt aquifer; aquifer microbes that can increase calcite precipitation; and, a heretofore unrecognized archaeal component in an oxic aquifer.

Future goals aim to quantify the volumetric productivity of subsurface microorganisms that are degrading chlorinated hydrocarbons, identify unique messenger RNA that signifies bioremedial activities, and directly detect microorganisms on minerals using secondary ion mass spectroscopy.

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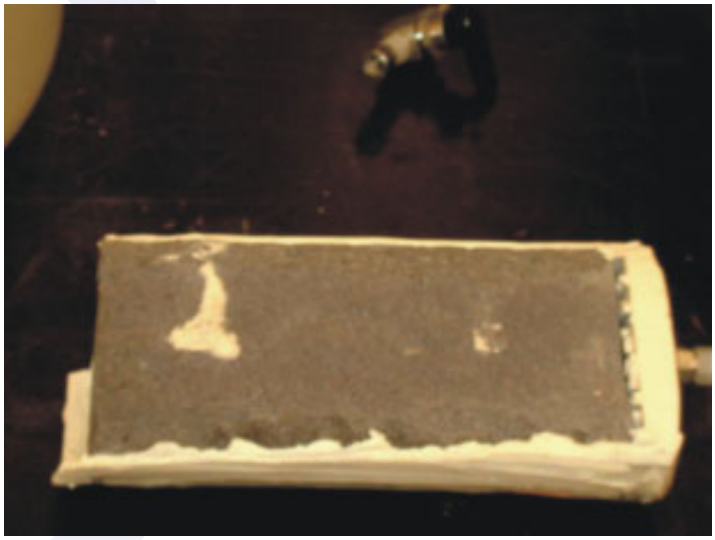

Science

INL

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## Selected Publications/ Presentations/Patents

**F.S. Colwell and R.M. Lehman,** "Carbon Source Utilization Profiles for Microbial Communities from Hydrologically Distinct Zones in a Basalt Aquifer," *Microbial Ecology*, Vol. 33, 1997, pp. 240–251.

**F.S. Colwell, T.C. Onstott, M.E. Delwiche, D. Chandler, J.K. Fredrickson, Q.J. Yao, J.P. McKinley, D.R. Boone, R. Griffiths, T.J. Phelps, D. Ringelberg, D.C. White, L. LaFreniere, D. Balkwill, R.M. Lehman, J. Konisky, and P.E. Long,** "Microorganisms from Deep, High Temperature Sandstones: Constraints on Microbial Colonization," *FEMS Microbiology Reviews*, Vol. 20, 1997, pp. 425–435.

**T.C. Onstott, T.J. Phelps, F.S. Colwell, D. Ringelberg, D.C. White, D.R. Boone, J.P. McKinley, T.O. Stevens, P.E. Long, D.L. Balkwill, W.T. Griffin, and T. Kieft,** "Observations Pertaining to the Origin and Ecology of Microorganisms Recovered from the Deep Subsurface of Taylorsville Basin, Virginia," *Geomicrobiology*, Vol. 15, 1998, pp. 353–385.

**D.E. Cummings, S. Spring, and R.F. Rosenzweig,** "The Ecology of Iron-Reducing Bacteria in Contaminated and Pristine Environments," In *Manual of Environmental Microbiology,* 2nd edition, Hurst, C. J., R. L. Crawford, G. R. Knudsen, M. J. McInerney, and L. D. Stetzenbach, eds., ASM Press, Washington, DC, 2002.

**D.E. Cummings, A.W. March, B. Bostick, S. Spring, F. Caccavo, Jr., S. Fendorf, and R.F. Rosenzweig**. "Evidence for Microbial Fe(III) Reduction in Anoxic, Mining-Impacted Lake Sediments (Lake Coeur d'Alene, Idaho)," Appl. Environ. Microbiol. Vol. 66, 2000, pp.154-162.

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**R.M. Lehman, F.F. Roberto, D. Early, D.F. Bruhn, S.E. Brink, S.P. O'Connell, M.E. Delwiche and F. Colwell**, "Attached and Unattached Microbial Communities in Closely-Paired Groundwater and Core Samples From an Acidic, Crystalline Rock Aquifer," *Appl. Environ. Microbiol*. Vol. 67, 2001, pp. 2095-2106.

**R.M. Lehman, F.S. Colwell, and G.A. Bala**, "Attached and Unattached Microbial Communities in a Simulated Basalt Aquifer Under Fracture- and Porous-Flow Conditions," *Appl. Environ. Microbiol*. Vol. 67, 2001, pp. 2799-2809.